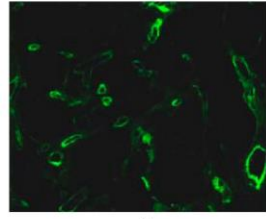
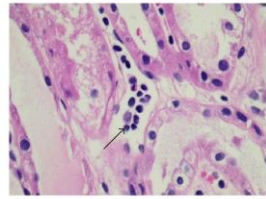
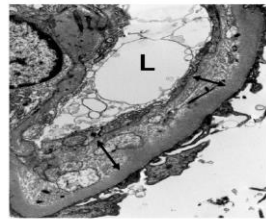
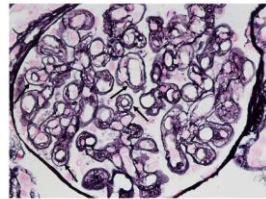
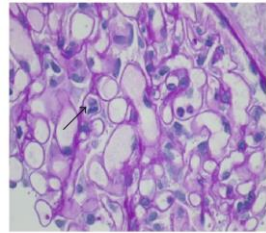
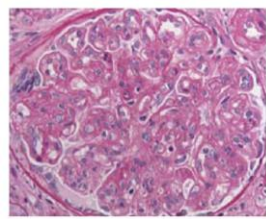


Chronic Antibody Mediated Rejection (cAMR)



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PROFESSOR OF MEDICINE, HASHEMI NEJAD KIDNEY HOSPITAL

IRAN UNIVERSITY OF MEDICAL SCIENCES

Introduction

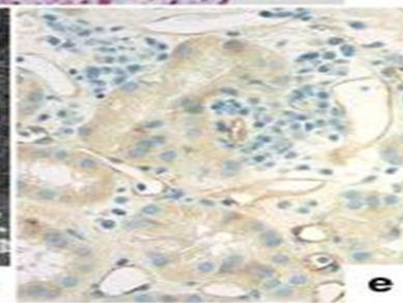
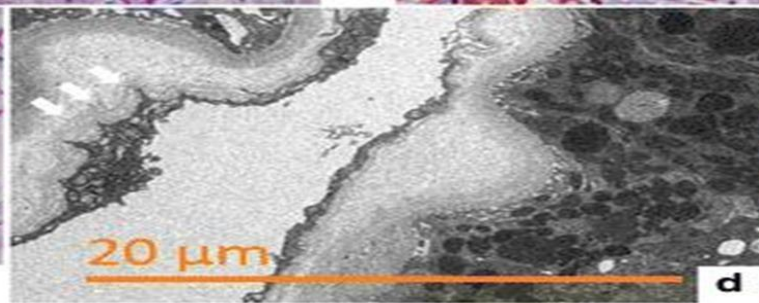
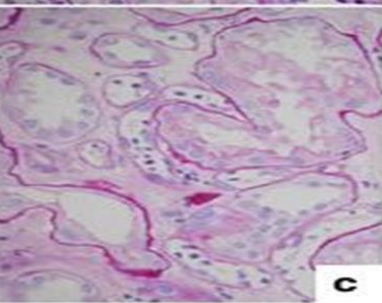
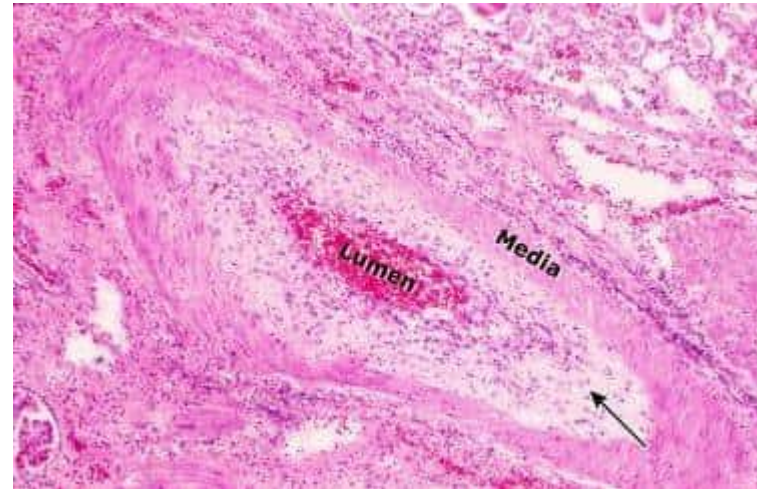
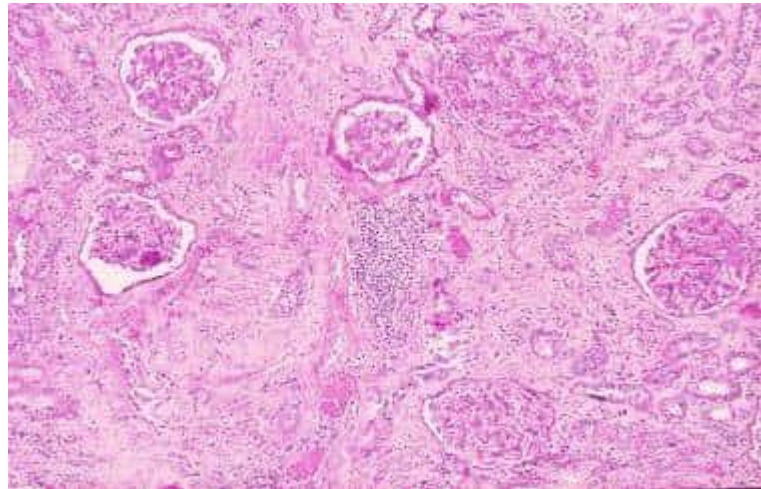
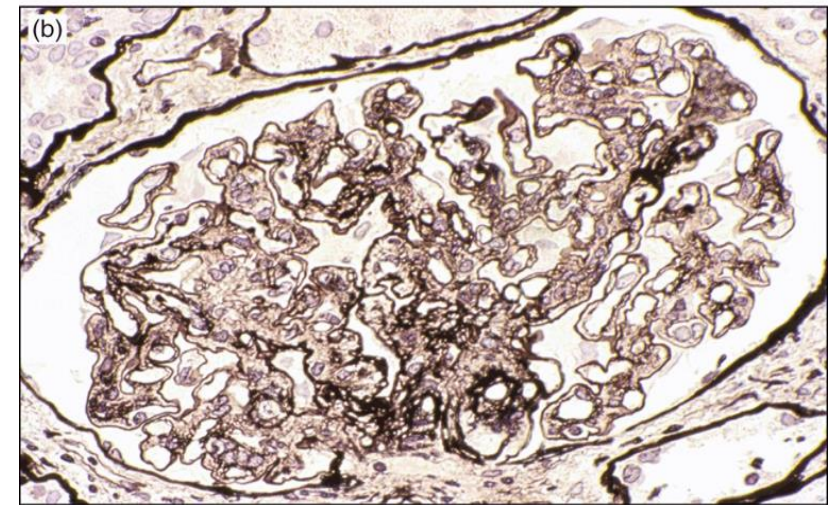
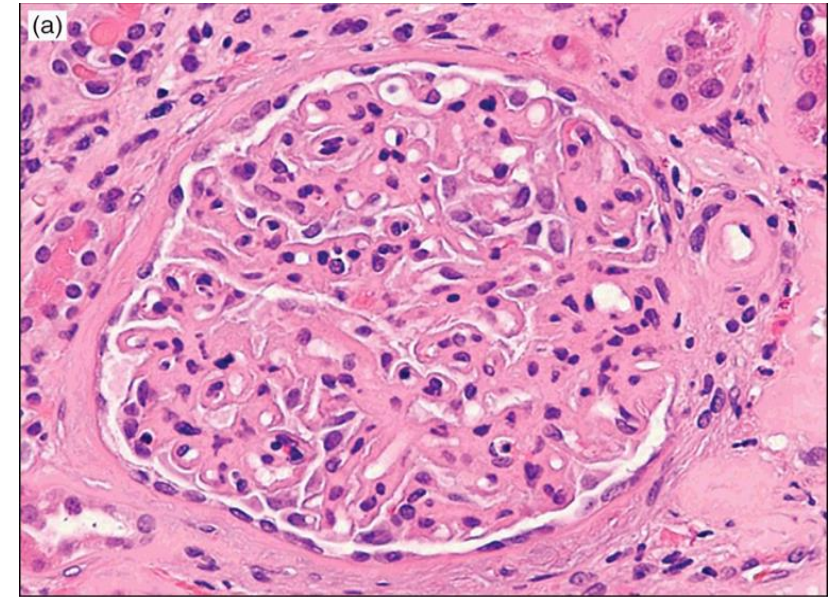
cAMR is the most important cause of late kidney allograft failure and a major unmet therapeutic challenge.

Detected in approximately 20–30% of kidney transplant recipients over time and is one of the leading causes of late graft dysfunction.

Up to 40–60% of patients with cAMR progress to graft loss within 3–5 years, especially high-risk group with persistent high-titer DSA and late diagnosis

Definition cAMR : Evidence of chronic tissue injury

Transplant glomerulopathy (cg > 0)
Severe peritubular capillary basement membrane Multilayering
Interstitial fibrosis / Tubular atrophy (IFTA)±
Arterial intimal fibrosis (chronic vasculopathy)



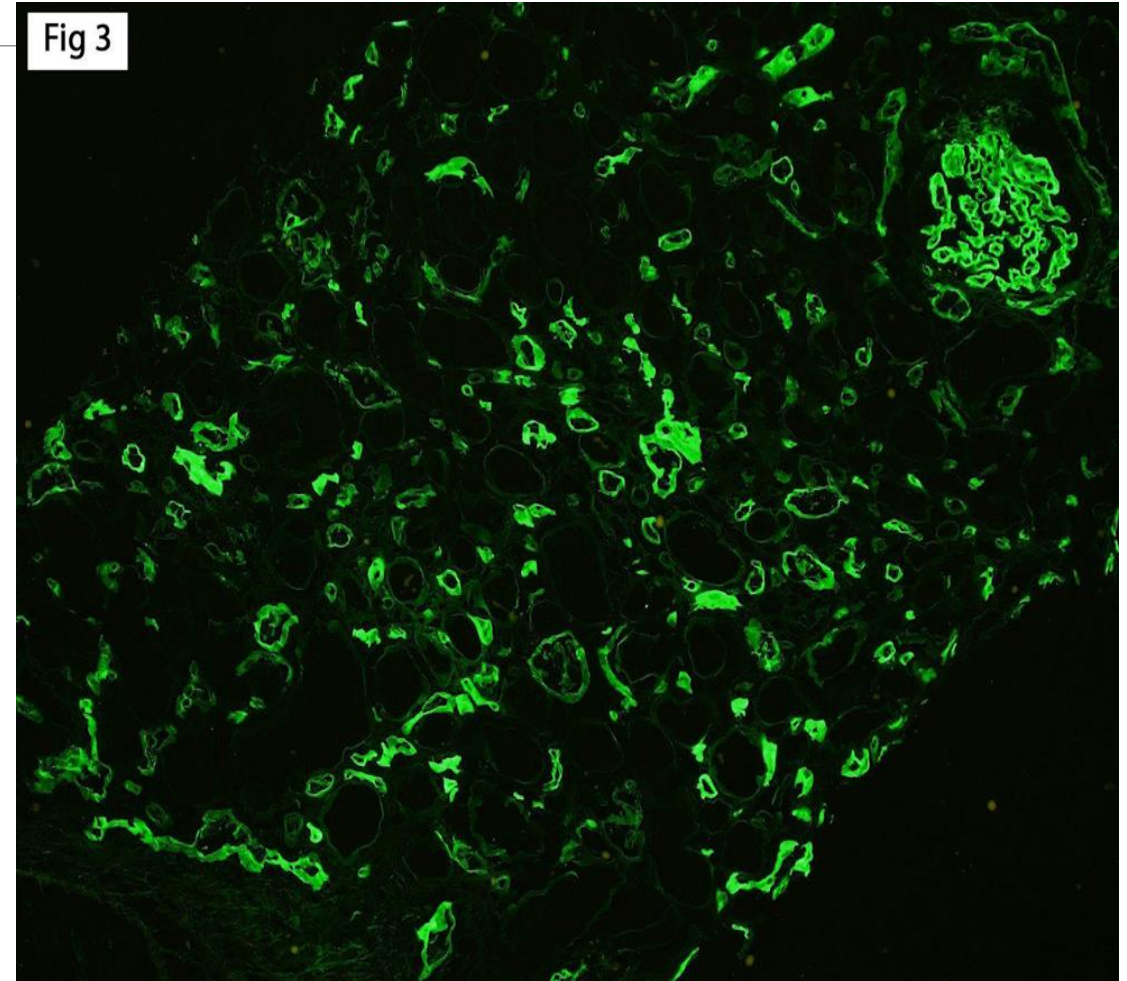
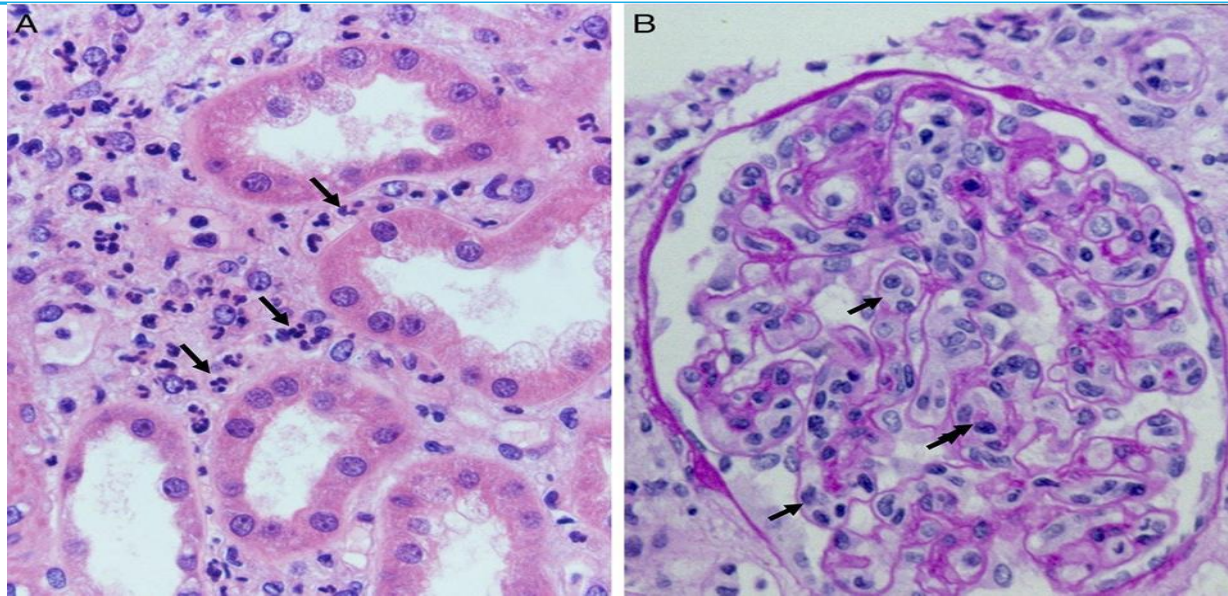
Definition cAMR : Evidence of Antibody–Endothelium Interaction

At least ONE of the following:

C4d deposition in peritubular capillaries

Microvascular inflammation (MVI): Glomerulitis (g) Peritubular capillaritis (ptc)

Molecular markers of endothelial activation (ENDATs / transcriptomics)



Molecular markers of endothelial activation (ENDATs / transcriptomics)

Endothelial activation & adhesion: VCAM1 (Vascular Cell Adhesion Molecule-1), ICAM1 (Intercellular Adhesion Molecule-1), E-selectin

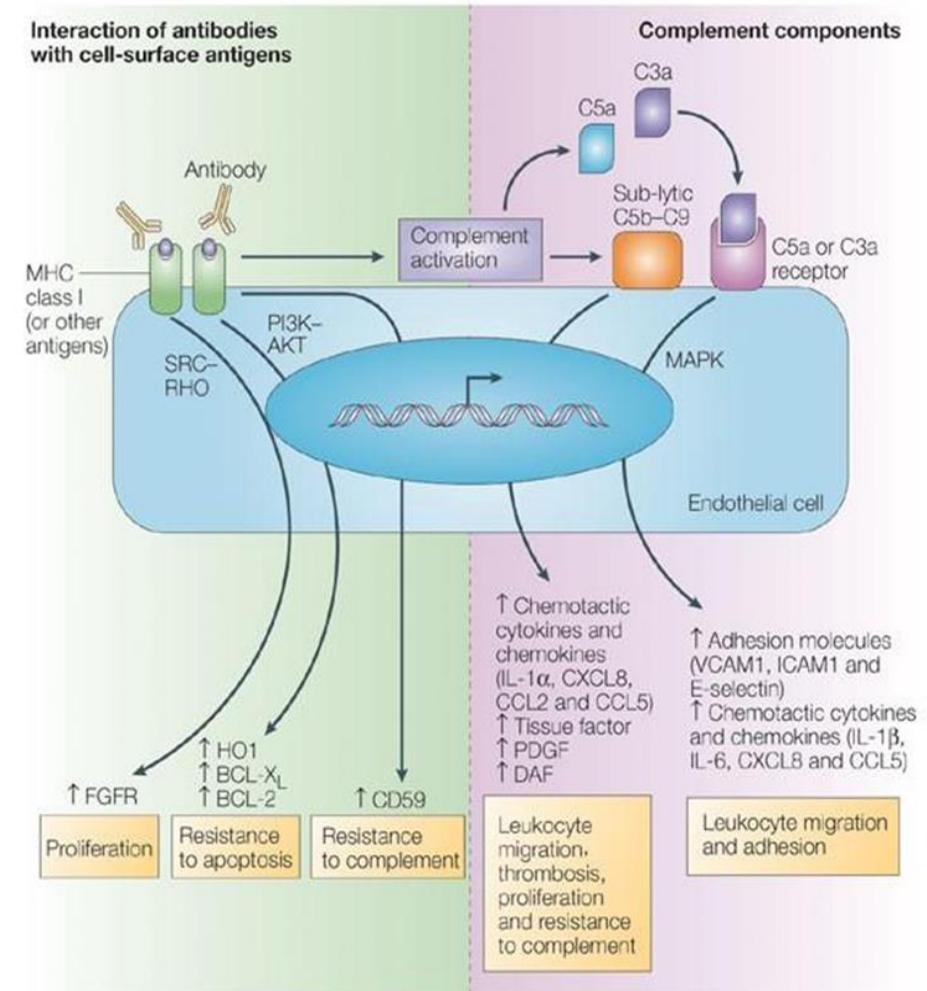
Interferon signaling pathway: STAT1, IRF1

Interferon- γ -inducible chemokines : CXCL9, CXCL10 (IP-10), CXCL11

NK cell / cytotoxicity-associated transcripts : GNLY

(Granulysin), GZMB (Granzyme B), PRF1 (Perforin)

Endothelial injury / activation markers : VWF (von Willebrand factor), PECAM1 (CD31), CDH5 (VE-cadherin)



ENDAT Workflow: From Biopsy to Clinical Diagnosis

- ❓ Kidney Allograft Biopsy
- ❓ Tissue Allocation (Histology + Molecular core)
- ❓ RNA Extraction
- ❓ Gene Expression Profiling (CXCL9, CXCL10, VCAM1, STAT1)
- ❓ Bioinformatics Analysis
- 🏢 ENDAT Score + AMR Probability
- ❓ Integrated Interpretation (Banff + DSA)
- 🛡️ Final Diagnosis (e.g., Chronic Active AMR)

PATIENT INFORMATION

Patient ID: TXP-123456
Name: John Doe
DOB / Sex: 08/12/1972 (M)
Transplant Date: 11/15/2020
Time Post-Transplant: 3.5 years
Indication: Rising creatinine, suspected rejection



SPECIMEN INFORMATION

Specimen Type: Kidney Allograft Biopsy
Collection Date: 05/01/2024
Received Date: 05/02/2024
Specimen Number: S24-98765
Requesting Physician: Dr. A. Smith
Center: University Transplant Center



TEST PERFORMED

**Molecular Microscope
for Endothelial Activation
& Rejection (MMDx®)**

Method: RNA expression profiling
(Microarray)

RESULT SUMMARY

PROBABILITY OF ANTIBODY-MEDIATED REJECTION

**82%
HIGH**



Interpretation: High probability of antibody-mediated rejection.

ENDOTHELIAL ACTIVATION (ENDAT) SCORE



HIGH

Interpretation: Increased endothelial activation consistent with antibody-endothelium interaction.

OTHER MOLECULAR SCORES

	Score	Interpretation
TCMR Score	22%	LOW
Injury Score	67%	MODERATE-HIGH
Fibrosis / Atrophy Score	46%	MODERATE
Microvascular Inflammation Score	78%	HIGH

KEY ENDOTHELIAL ACTIVATION TRANSCRIPTS

Gene Group / Pathway	Representative Genes	Result
IFN-γ Inducible Chemokines	CXCL9, CXCL10, CXCL11	↑↑↑
Adhesion Molecules	VCAM1, ICAM1, SELE	↑↑↑
Cytotoxic / NK Cell Pathway	GZMB, PRF1, GNLY	↑↑
Endothelial Activation Markers	VWF, PECAM1, CDH5	↑↑
IFN-γ Signaling Pathway	STAT1, IRF1	↑↑

RESULT KEY

↑↑↑ Markedly Increased ↔ No Significant Change
↑↑ Moderately Increased ↓ Decreased
↑ Mildly Increased

MOLECULAR DIAGNOSTIC INTERPRETATION

The molecular profile demonstrates a high probability of antibody-mediated rejection with significant endothelial activation (ENDAT high). There is upregulation of IFN-γ-inducible chemokines and adhesion molecules, along with cytotoxic/NK cell-associated transcripts, consistent with active antibody-endothelium interaction and microvascular injury.

These findings support the diagnosis of antibody-mediated rejection in the appropriate histologic and clinical context.

RECOMMENDED CORRELATION

- Correlate with histology (Banff scores: g, ptc, cg, ptcml, ci, ct)
- Correlate with donor-specific antibody (DSA) testing (HLA ± non-HLA)
- Correlate with clinical findings and graft function
- Consider C4d staining results

OVERALL MOLECULAR IMPRESSION



**Consistent with
Chronic Active
Antibody-Mediated Rejection
(cAMR)**

Confidence: HIGH

Practical Use of ENDAT in the Clinic

POSITIVE POINTS

1. Clarifying “uncertain” biopsies

When histology is borderline (e.g., mild **g/ptc**, unclear chronicity), When **C4d is negative**

2. Diagnosing C4d-negative AMR

A common real-world problem, ENDAT can act as a **surrogate for antibody activity**

3. DSA-negative but suspicious cases

4. Risk stratification / prognosis

5. Complementing Banff (not replacing it)

LIMITATIONS

1. Lack of specificity

ENDAT reflects **inflammation**, not only AMR ,
Can be elevated in

- TCMR
- Viral infections (e.g., BK)
- Ischemic injury

2. No universal cut-off

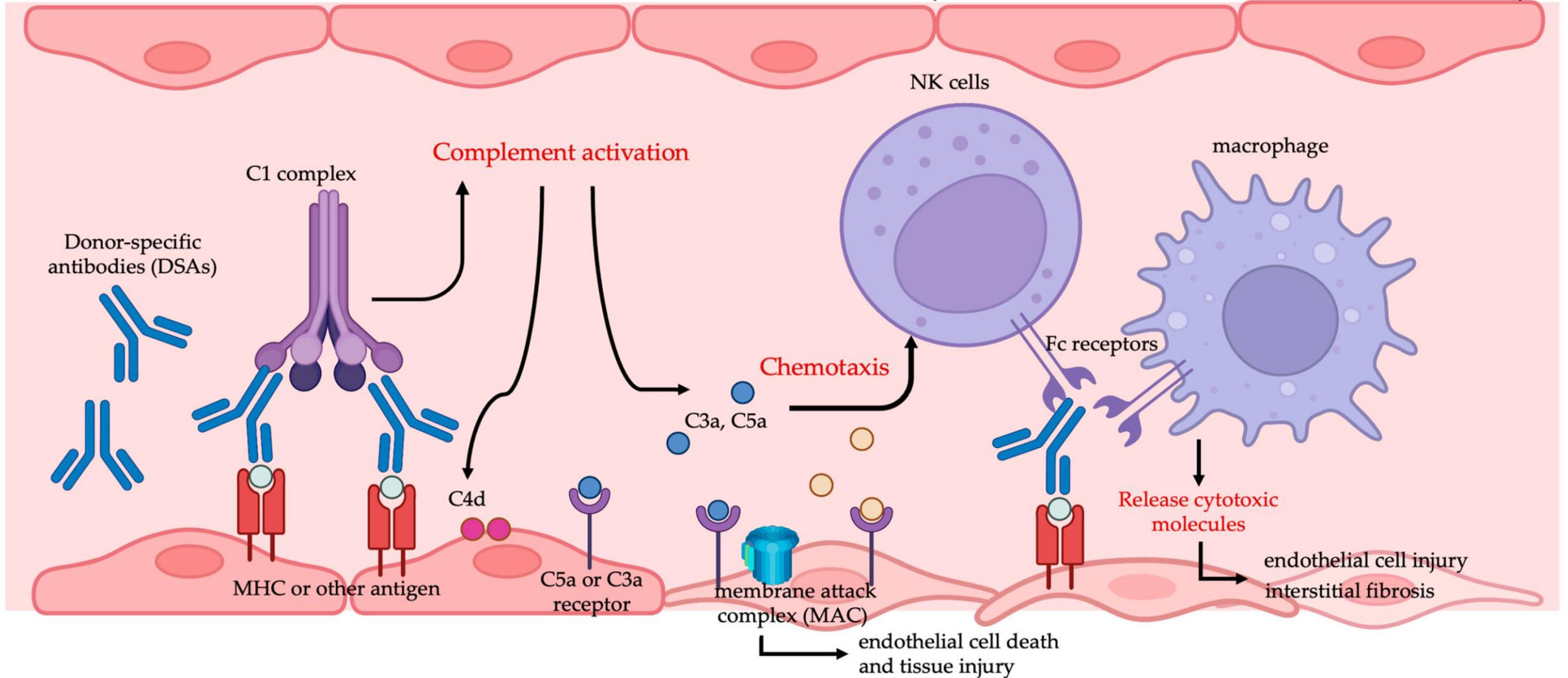
3. Limited availability & cost

4. Not yet fully guideline-integrated

5. Unclear role in therapy monitoring

Complement-dependent cytotoxicity

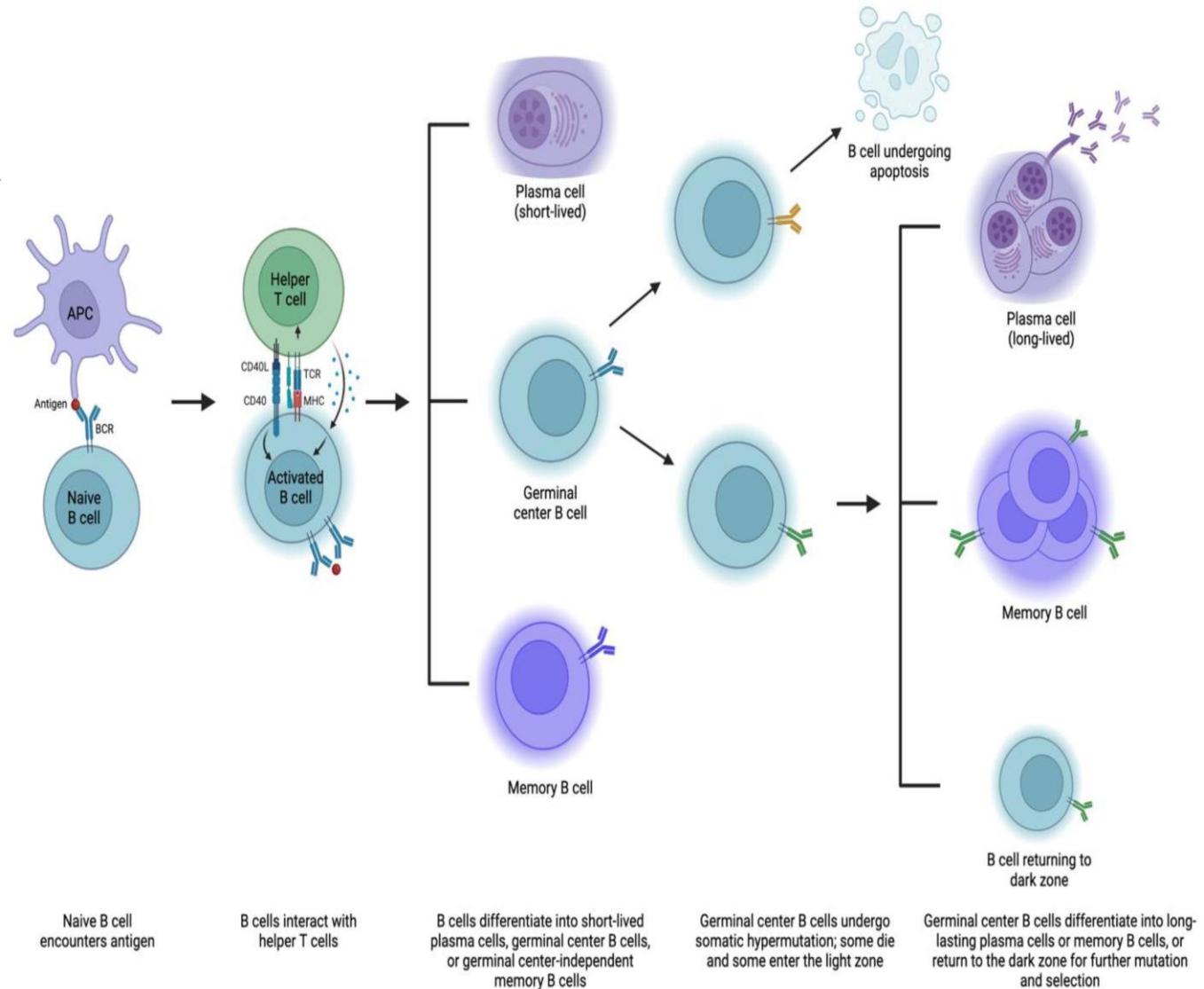
Complement-independent pathways of antibody-mediated cellular cytotoxicity



Hidden Players In Chronic Antibody-mediated Rejection (cAMR): Explaining Therapeutic Resistance”

- **Long-lived plasma cells (LLPCs)** → Continuous DSA production, resistant to conventional therapy
- **Memory B cells** → Rapid reactivation and regeneration of antibody response
- **T follicular helper (Tfh) cells** → Sustain B-cell maturation and antibody production
- **Natural killer (NK) cells** → Mediate antibody-dependent cellular cytotoxicity (ADCC)
- **Non-HLA antibodies** → e.g., anti-AT1R, anti-ETAR → complement-independent injury
- **Endothelial-to-mesenchymal transition (EndMT)** → Link between immune injury and chronic fibrosis
- **Innate immune activation (macrophages, monocytes)** → Amplification of microvascular inflammation
- **Extracellular vesicles & microRNAs** → Intercellular signaling, propagation of injury

In (cAMR), the “memory” of the immune system is a central driver of persistent and treatment-resistant injury. Two main cellular players are involved: memory B cells and long-lived plasma cells, both shaped by prior antigen exposure (donor HLA or non-HLA).



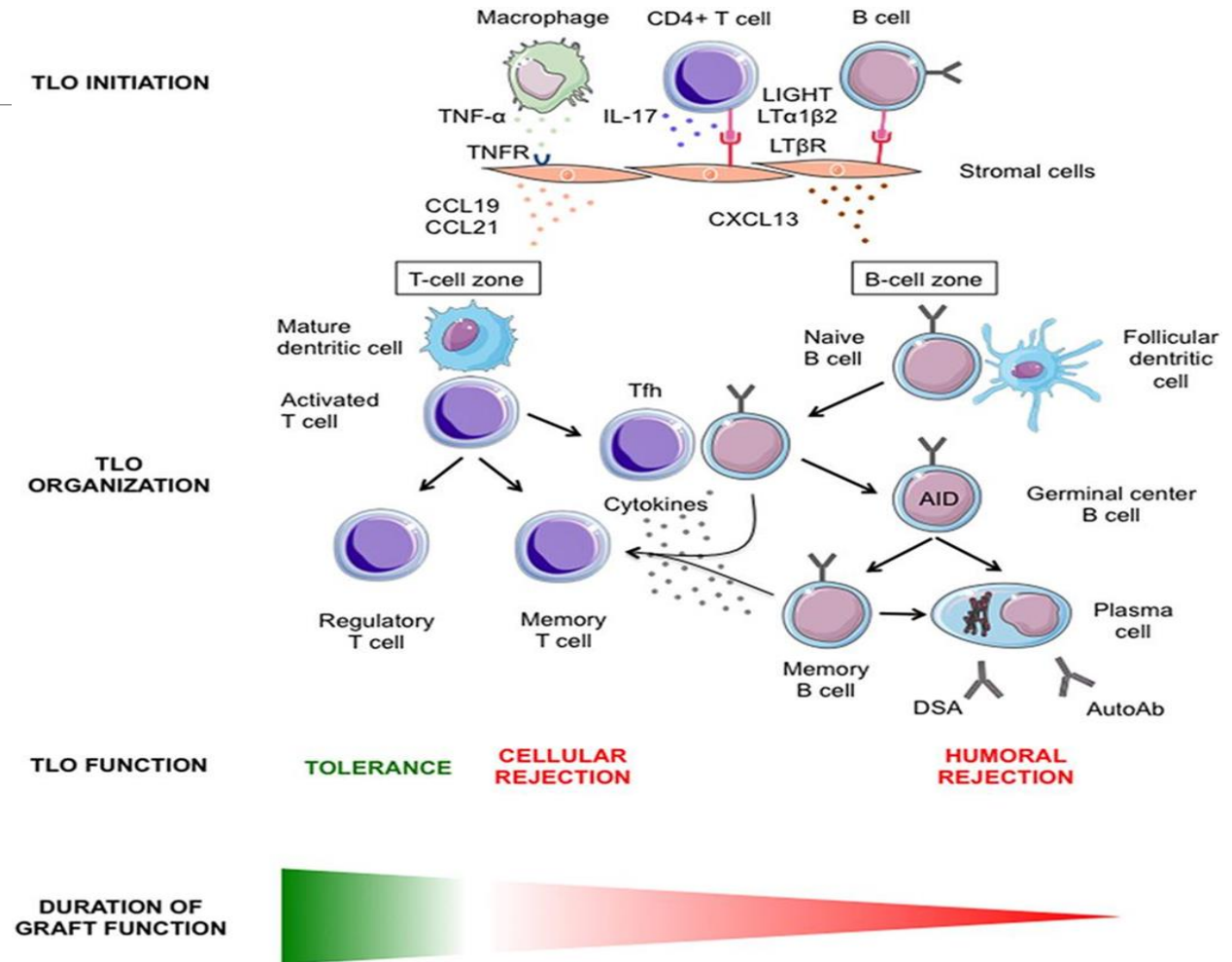
Memory B cells - the hidden reservoir

After initial sensitization (transplant, transfusion, pregnancy), Memory B cells specific for donor antigens persist long-term.

They are usually quiescent, not detected in routine assays, not producing antibodies continuously.

But **when reactivated**, rapidly differentiate into plasma cells and cause **a rapid rise in (DSA)**, drive progression of cAMR even years after transplant.

Key point: They explain why DSA can reappear suddenly after being negative



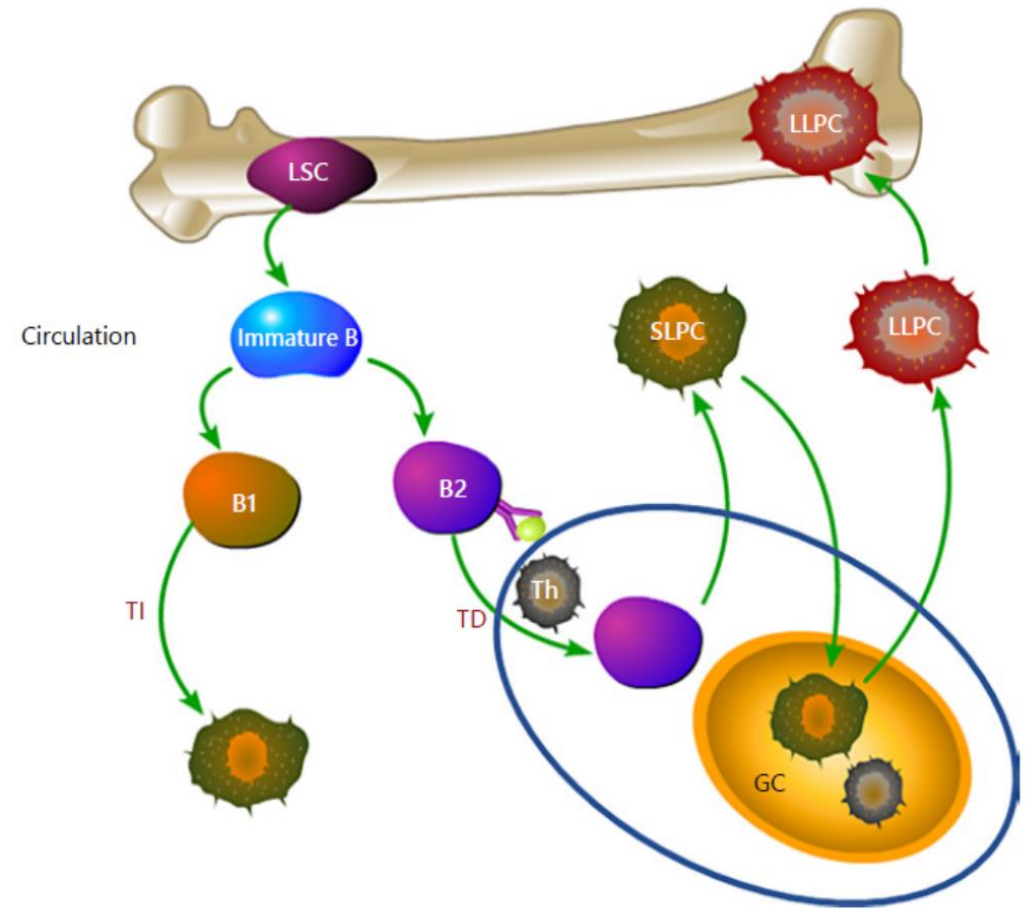
Long-lived plasma cells (LLPCs)

LLPC represent a minority of plasma cell pool (10-20%),
These cells are the major effector in cAMR: Reside in bone marrow and inflamed graft tissue, Continuously produce DSA (anti-HLA and non-HLA antibodies).

Do NOT require antigen stimulation once established.

Clinical importance: Resistant to standard therapies like:
Rituximab (targets CD20, but plasma cells are CD20-negative)

Maintain chronic endothelial injury



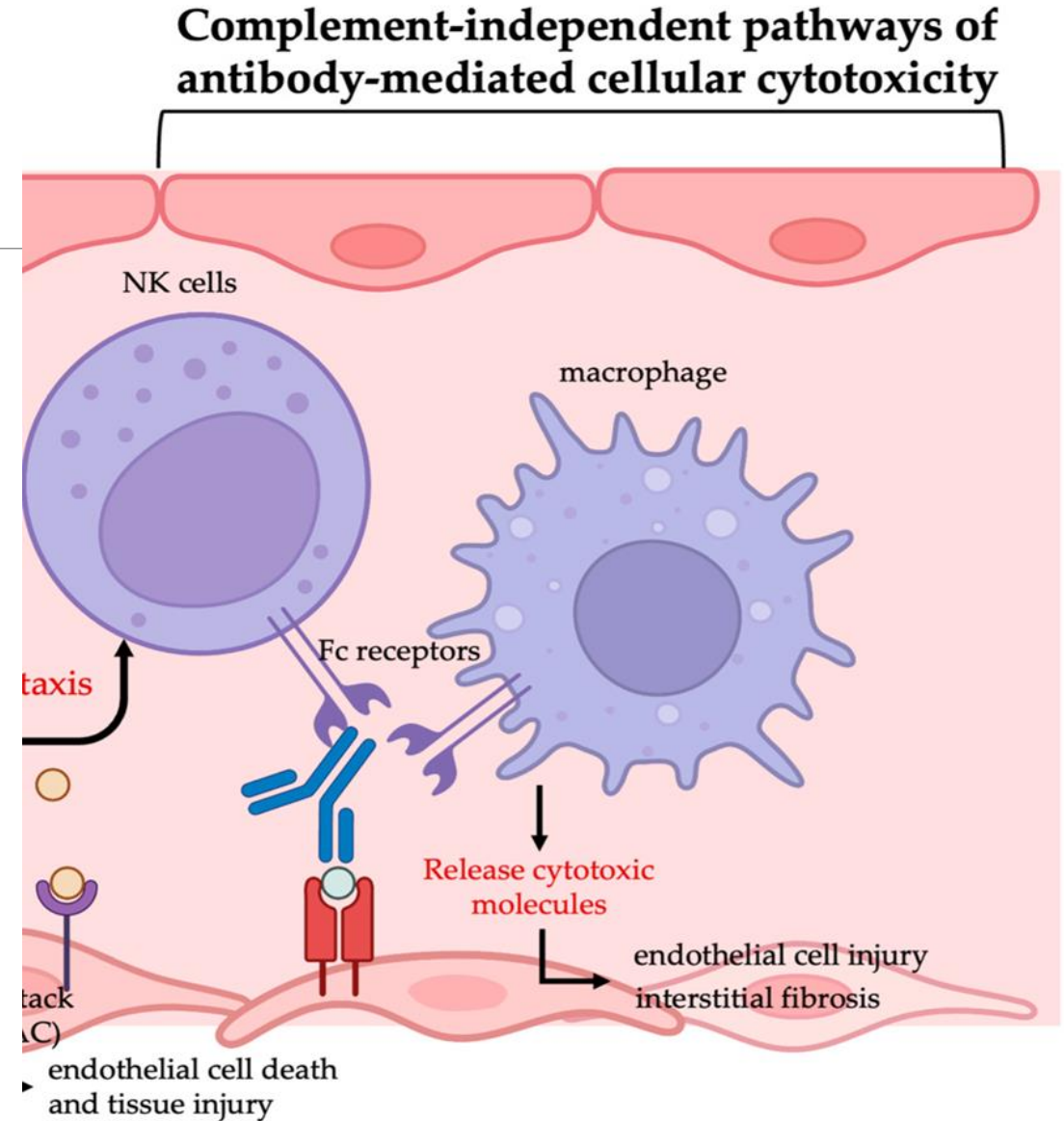
Natural killer (NK) cells

Natural killer (NK) cells :a key effector arm in cAMR—
sometimes even more important than complement.

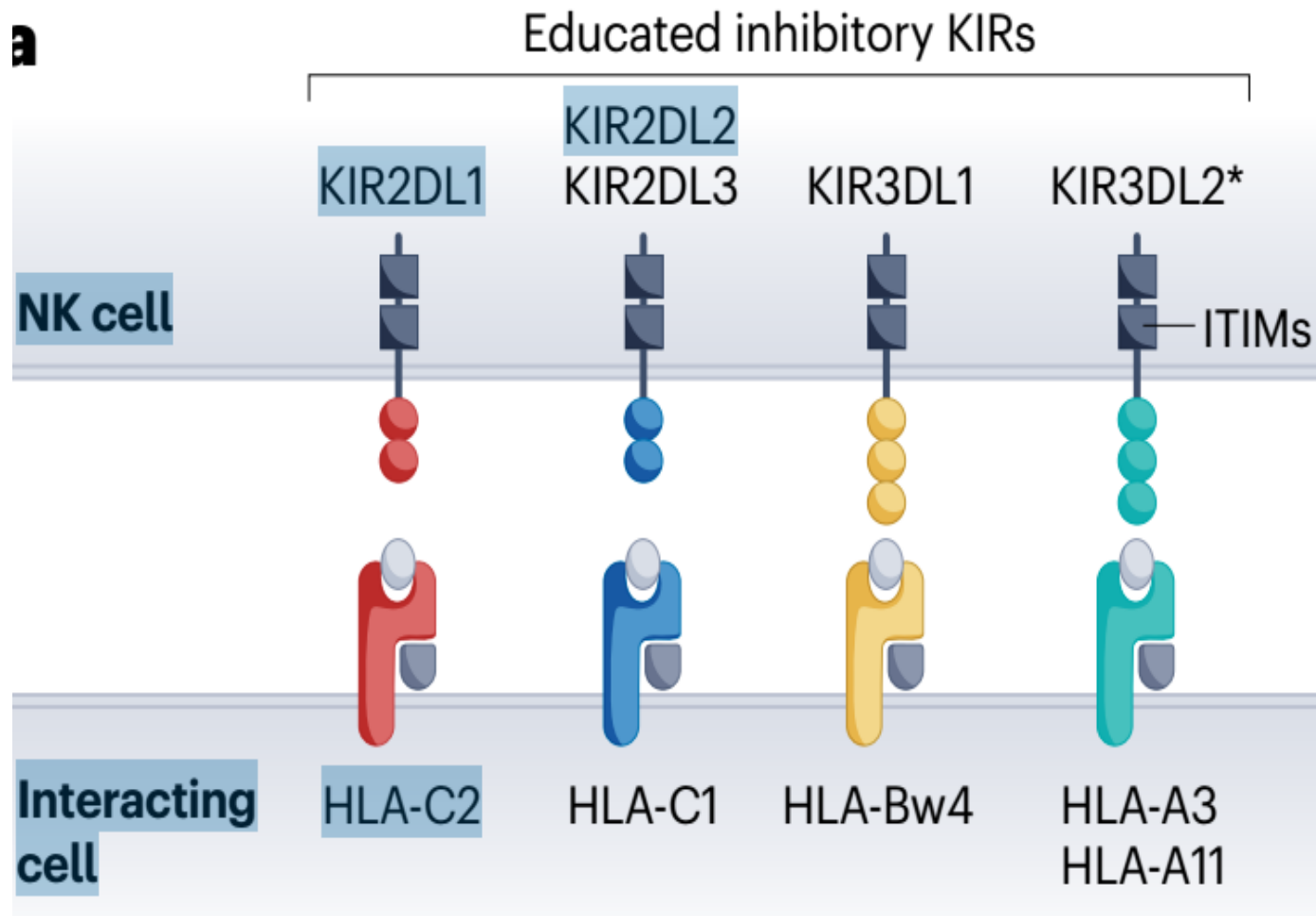
NK cells express CD16 (FcγRIII), which binds antibodies
already attached to graft endothelium.

Sequence: Donor-specific antibodies (DSA) bind endothelial
HLA (or non-HLA targets)NK cells attach via CD16, become
activated → release: Perforin + granzyme → endothelial cell
injury, IFN-γ → inflammation

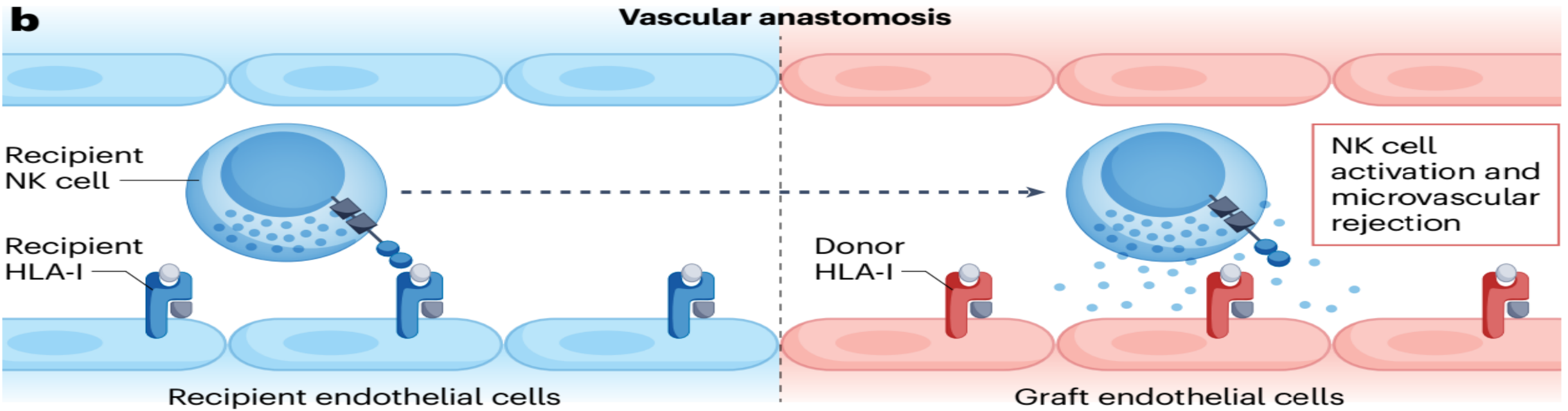
It is complement-independent



Missing-self-induced NK-mediated innate microvascular rejection



Schematic of the molecular interactions between **the five types of inhibitory killer immunoglobulin-like receptors (KIRs)** expressed on the surface of natural killer (NK) cells and the type I human leukocyte antigen (HLA) molecules expressed on the surface of interacting cells



HLA mismatches between donors and recipients create a situation in which graft endothelial cells are unable to provide HLA-I-mediated inhibitory signals to recipient circulating NK cells. This situation of missing self triggers activation of the recipient's NK cells, which initiates a pathophysiological cascade similar to antibody-mediated rejection (albeit in the absence of donor-specific antibodies).

Non HLA Antibody (C4d Negative , DSA negative)

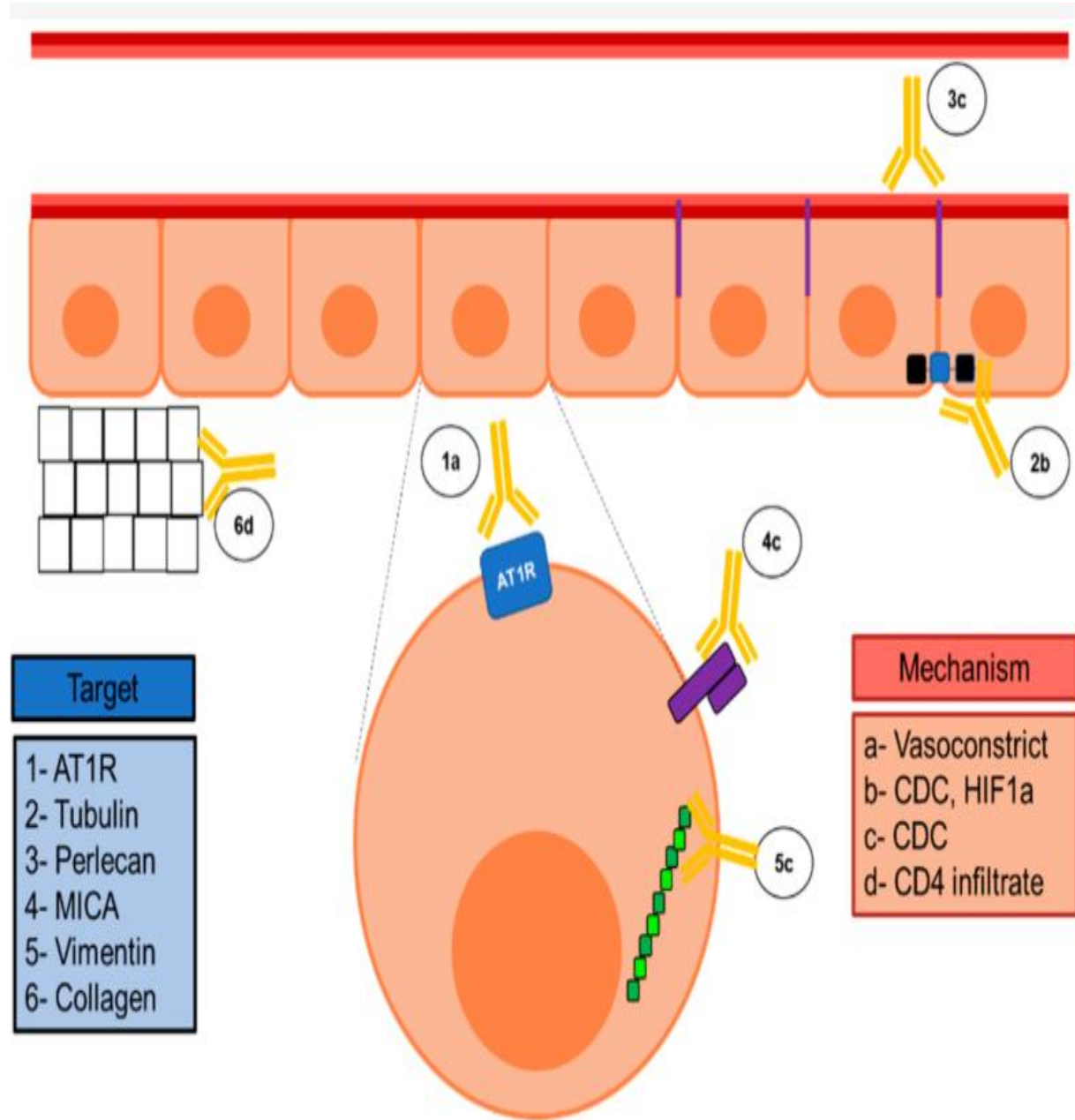
Terasaki deduced that non-HLA immunological factors contribute more to graft failure than HLA antibodies do (40% and 20%, respectively).

It is still debated whether these antibodies can act alone or whether they result in worse allograft outcome together with DSAs.

- Major histocompatibility complex class I-related chain A (MICA) and MICB
- Antibodies against G-coupled receptors present on the endothelium: AT1R and endothelin A receptor (ETAR).
- Antibodies to Collagen I, Collagen V, and k-alpha Tubulin
- Anti- perlecan/ LG3 fragment

1. Trigger antibody binds endothelial targets
2. Direct endothelial activation :Turn on intracellular pathways →Vasoconstriction, Inflammation (IL-6, MCP-1), Pro-fibrotic signaling
3. Immune amplification via NK cells
4. Complement (secondary, not dominant): Some non-HLA antibodies activate complement
5. Injury → autoimmunity loop

Final outcome in the graft :Persistent endothelial activation, Capillaritis +glomerulitis, Transplant glomerulopathy and Progressive fibrosis



Practical clinical insight

Think of non-HLA antibodies when you see:

C4d-negative AMR

DSA-negative rejection

Disproportionate vascular injury

Progressive fibrosis despite treatment

IL-6 in Chronic Antibody-Mediated Rejection (CAMR)

Central Role of IL-6

Produced by endothelial cells, macrophages, and lymphocytes, Links innate immunity to humoral alloimmunity, Promotes Tfh expansion and plasma cell survival.

Sustains donor-specific antibody (DSA) production and amplifies endothelial inflammation and fibrosis

IL-6–Driven CAMR Cascade

DSA → Endothelial Injury

Macrophage Activation

↑ IL-6 Release

↑ Tfh + Plasma Cells

Persistent DSA Production

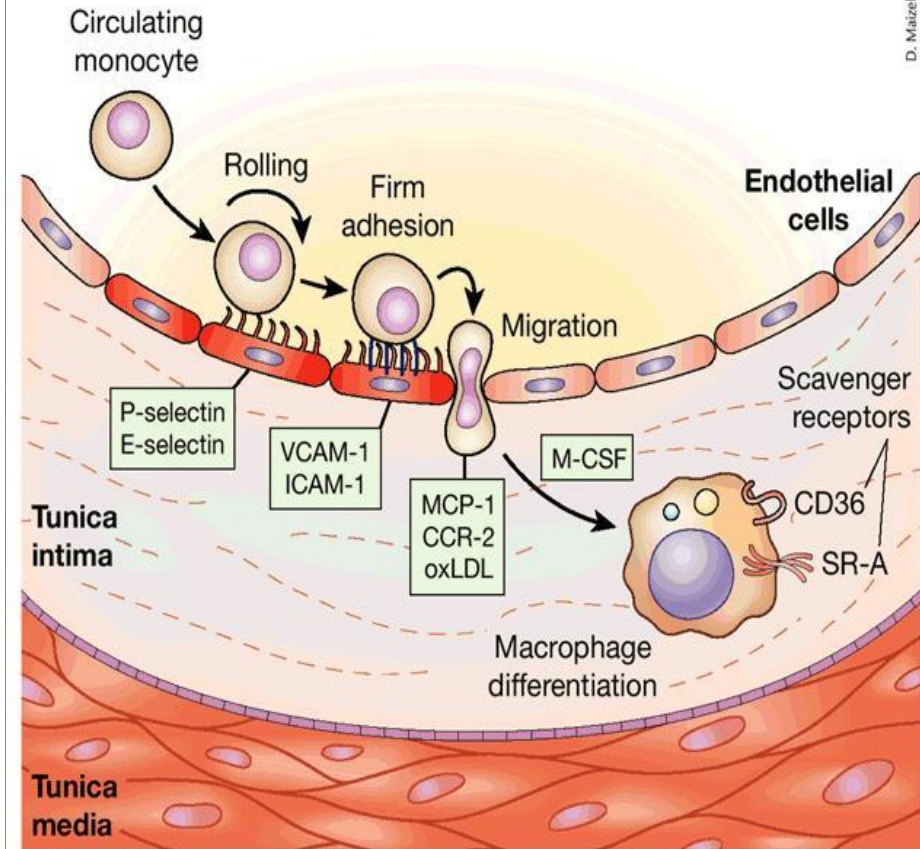
Microvascular Inflammation

Fibrosis & Transplant Glomerulopathy

Key Concept: IL-6 acts as a bridge cytokine linking endothelial injury, innate immunity, humoral alloimmunity, and chronic graft fibrosis.

Innate immune activation (monocytes & macrophages) in cAMR

1. Recruitment
 2. Activation pathways
 - 3- Complement fragments
 - 4- Release: TNF- α , L-1, Reactive oxygen species, Sustains vascular damage
Bridge to fibrosis (critical role)
- Macrophages produce: TGF- β \rightarrow drives, EndoMT & fibrosis, PDGF \rightarrow fibroblast activation, Matrix components, Central to: Transplant glomerulopathy, Interstitial fibrosis



Macrophage polarization

M1 (pro-inflammatory)

Early phase

Cytotoxic, cytokine-rich

M2 (pro-fibrotic)

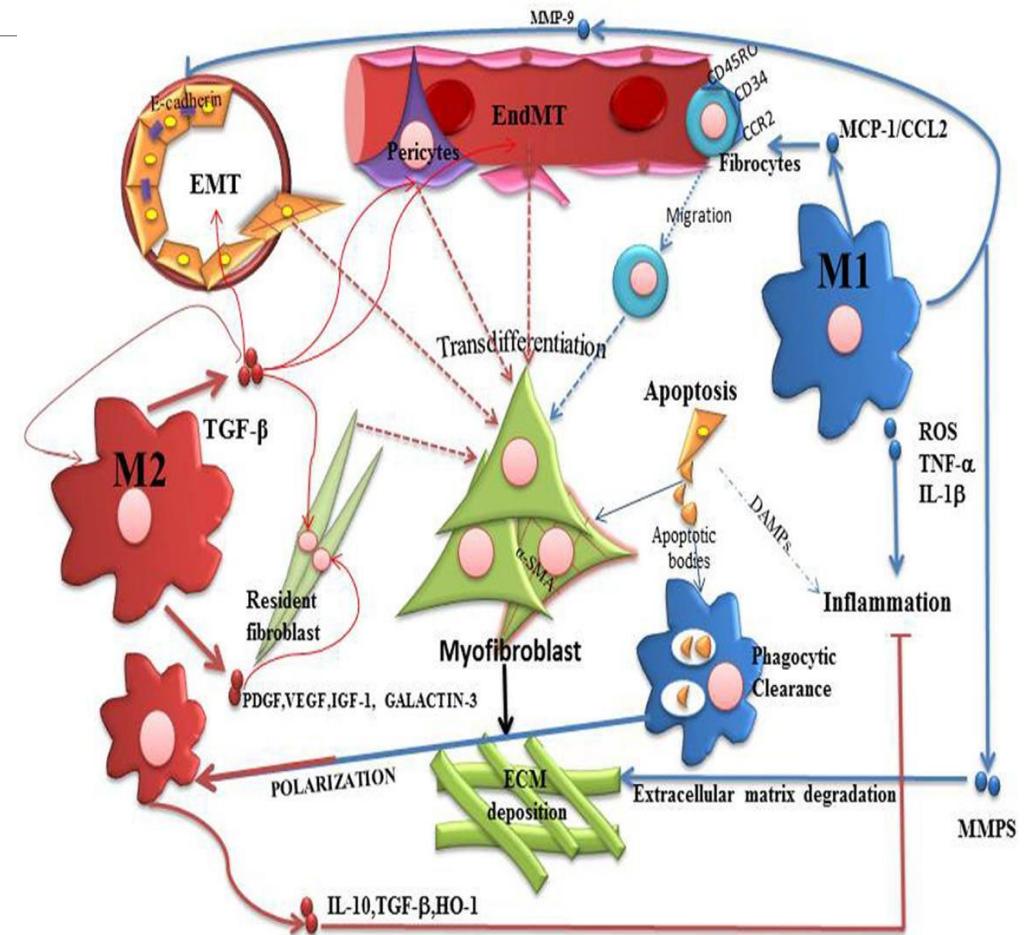
Chronic phase

Promote:

- Tissue remodeling
- Fibrosis

In cAMR:

Shift from M1 → M2 → chronic fibrotic phenotype



Tarcio Teodoro Braga et al, Molecular Innate Immunity 2015

Endothelial-to-mesenchymal transition (EndoMT) in cAMR

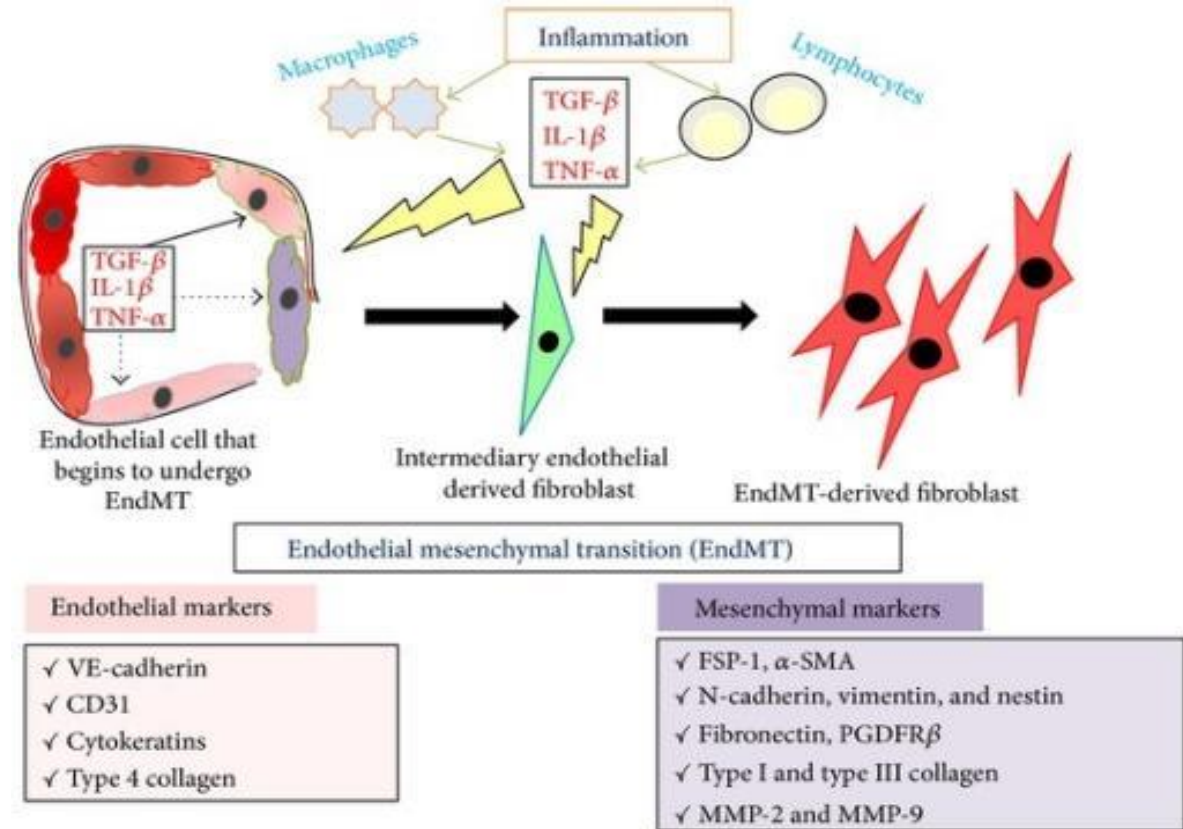
In chronic antibody-mediated rejection:

Persistent endothelial injury (HLA + non-HLA antibodies)

NK-cell-mediated damage

Cytokine-rich environment

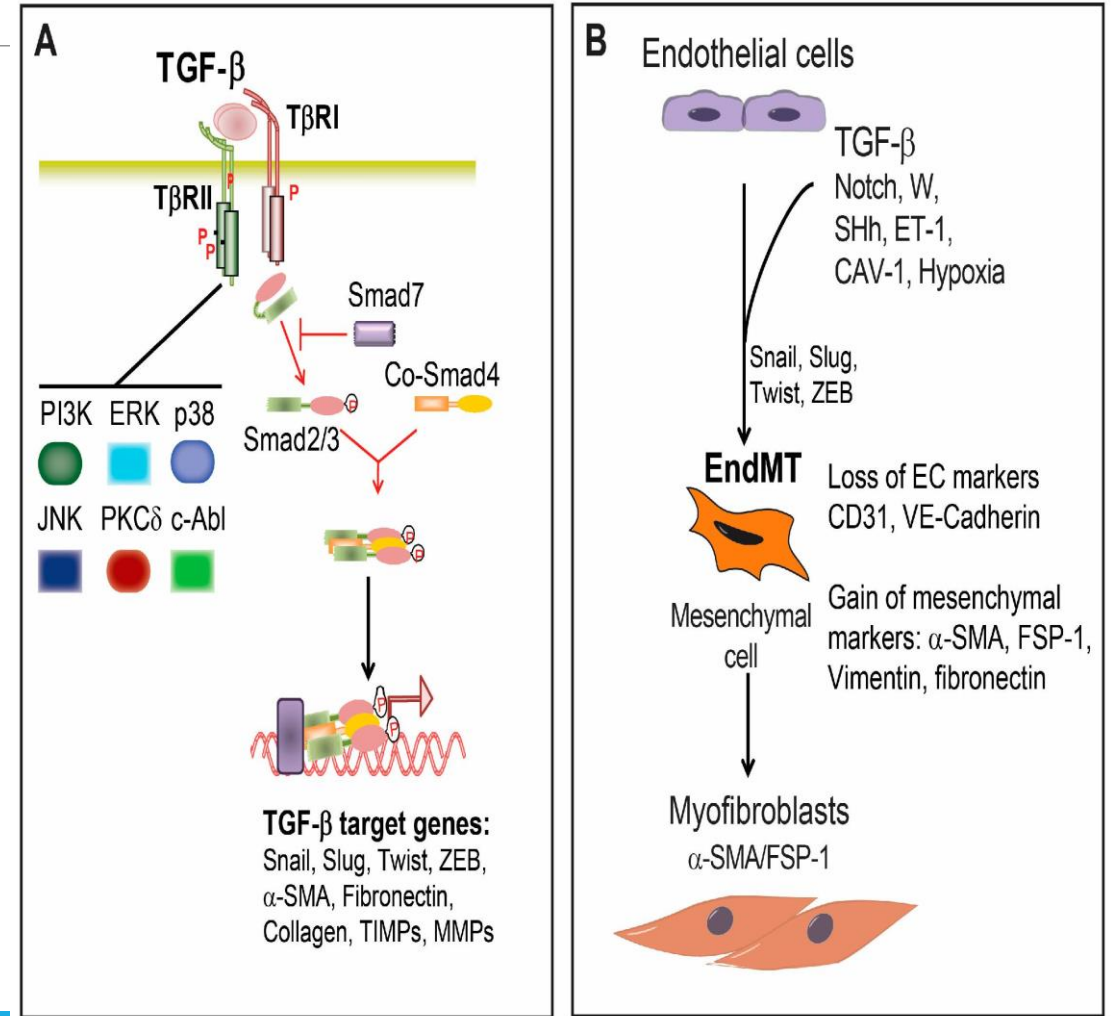
Drives **progressive EndoMT** → **fibrosis**



Mechanistic cascade

- Antibodies bind endothelium
- NK cells + inflammation injure vessels
- Endothelium becomes activated
- TGF- β signaling turns on
- Endothelial cells transition \rightarrow mesenchymal phenotype
- Fibroblast-like cells produce matrix
- Leads to: Transplant glomerulopathy, Interstitial fibrosis.

Evangelia Pardali, Int. J. Mol. Sci. 2017,



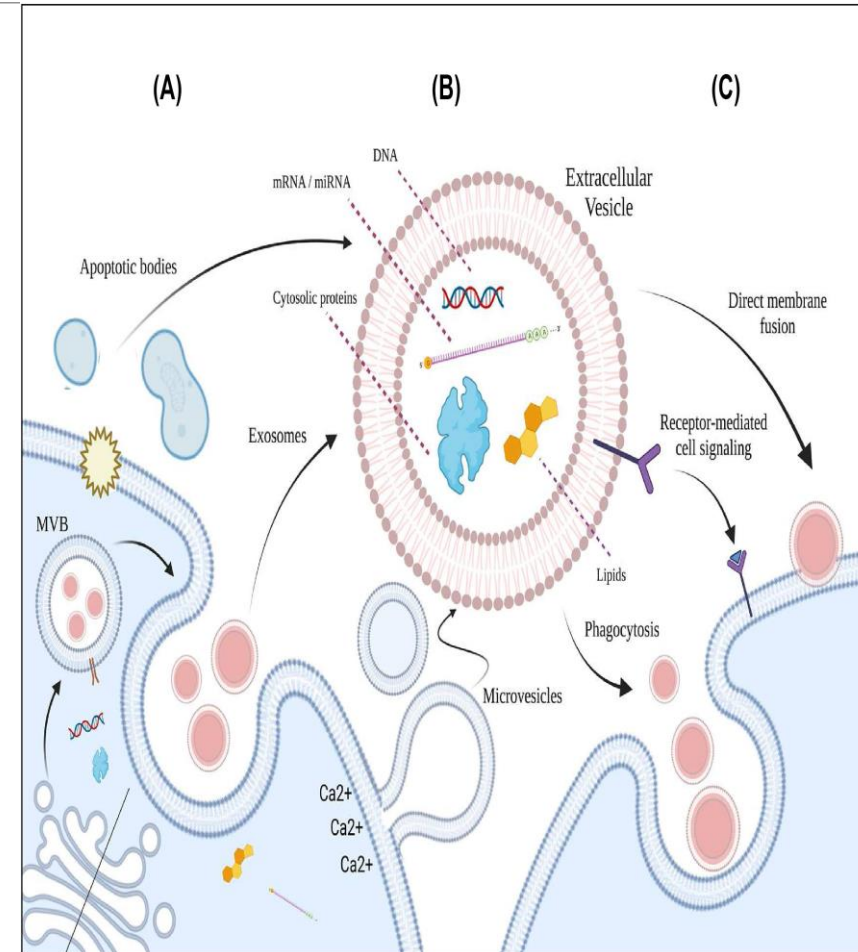
Extracellular vesicles (EVs) & microRNAs in cAMR

EVs (especially exosomes) are “message carriers” that shuttle microRNAs and proteins between endothelial cells and immune cells → amplifying inflammation and fibrosis in cAMR.

EVs sit in the middle of the network: Antibodies → endothelial injury, Endothelium → releases Evs, EVs → activate: NK cells, Macrophages, Fibrosis pathways, Creates a long-range communication system sustaining cAMR.

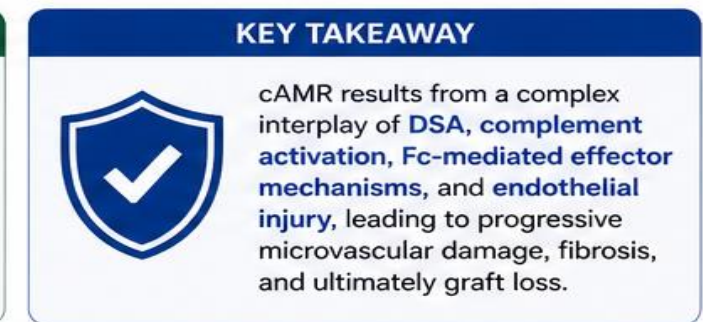
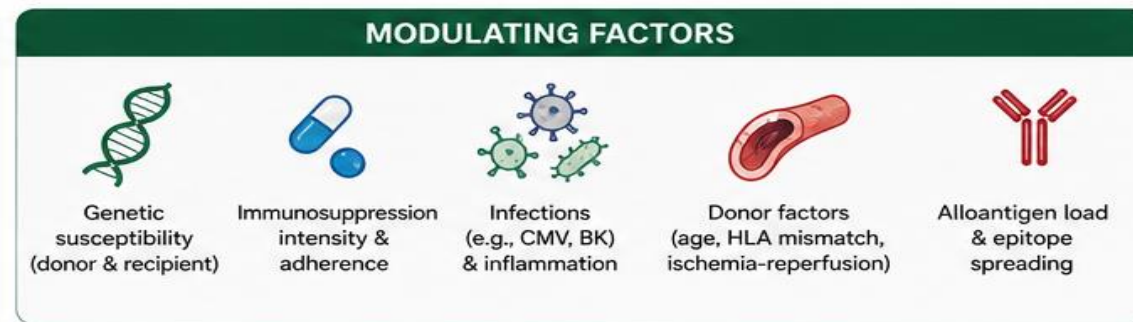
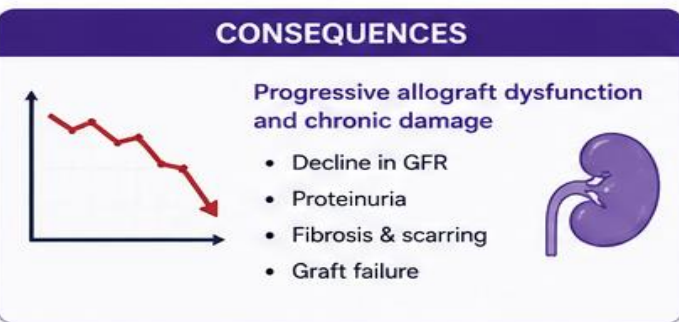
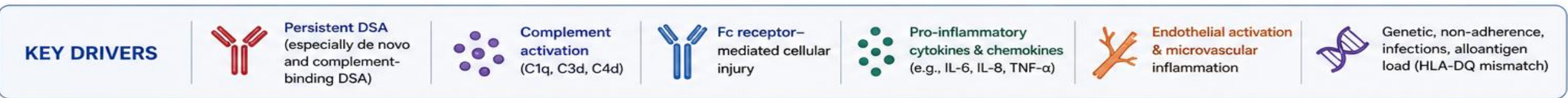
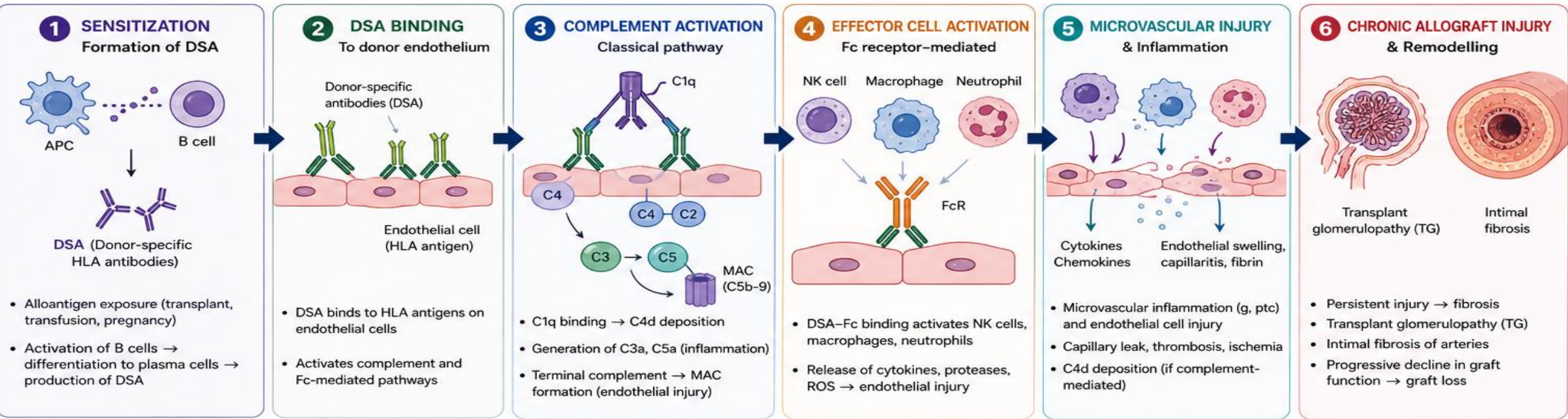
Clinical implications: Non-invasive biomarkers Blood or urine EV-miRNA profiles may detect cAMR early, monitor treatment response

Nassar et al , Front. Transplant., 2023



WRAP-UP: PATHOGENESIS OF cAMR

From sensitization to chronic allograft injury and loss



DSA in Chronic AMR – Decision Tree

When to Request & How to Monitor

WHEN TO REQUEST DSA

1. ROUTINE SURVEILLANCE (even if stable)

- Baseline (pre-transplant / immediate post-Tx)
- 3 months
- 6 months
- 12 months
- Annually thereafter



Goal: detect de novo DSA early
(before damage)

2. TRIGGER-BASED TESTING (Request immediately if)



GRAFT DYSFUNCTION

- Rising creatinine
- New or worsening proteinuria



IMMUNOLOGIC RISK EVENTS

- Non-adherence
- Reduction of immunosuppression
- Infection requiring ↓ immunosuppression



SENSITIZING EVENTS

- Blood transfusion
- Pregnancy
- Re-transplantation



BIOPSY FINDINGS

- Microvascular inflammation
- Early transplant glomerulopathy (TG)
- C4d ±

HOW TO INTERPRET DSA



STRONG RISK FEATURES

- De novo DSA
- Class II (especially DQ)
- High MFI
- Complement-binding (C1q+, C3d+)
- Persistent over time



LIMITATIONS

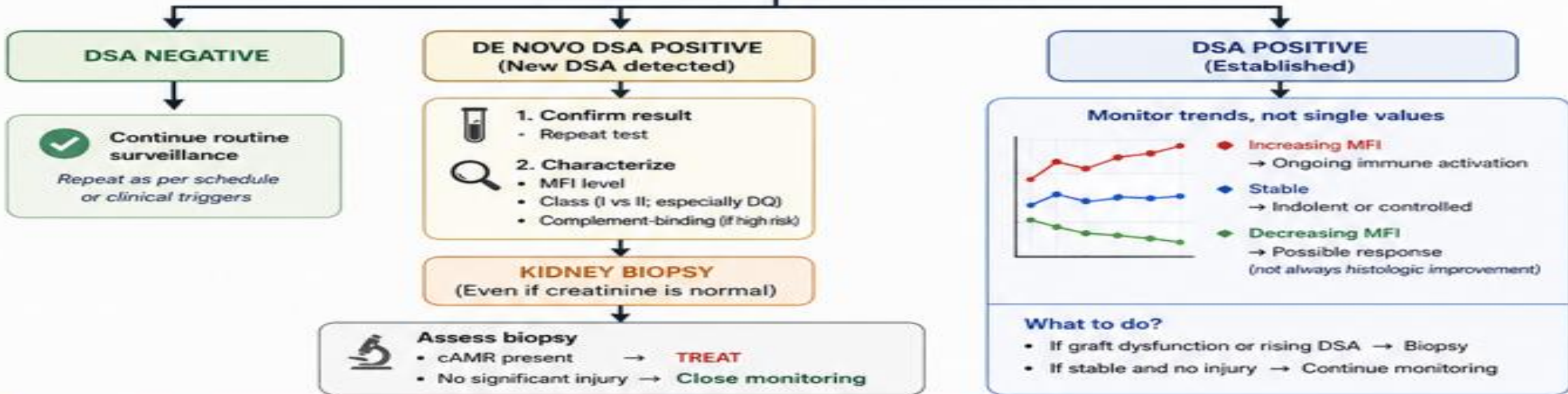
- DSA alone ≠ diagnosis
- Some DSA-positive patients have no injury (*accommodation*)
- Some cAMR cases are DSA-negative (non-HLA Ab)
- MFI is semi-quantitative, not absolute



PEARLS

- De novo DSA = strongest predictor of cAMR
- Class II (DQ) = most dangerous long-term
- Trend > absolute MFI
- DSA precedes histologic damage

DSA TEST RESULT



MONITORING FREQUENCY (After DSA is Positive)

HIGH-RISK DSA (De novo, high MFI, DQ, complement-binding, rapidly rising)	STABLE DSA (Low/Moderate risk, stable levels)	DSA NEGATIVE (Stable patient)
Every 1–3 months	Every 3–6 months	As per routine schedule

IMPORTANT PITFALLS

- Do NOT treat numbers alone (High MFI without injury ≠ automatic treatment)
- MFI is not absolute (Lab variability, prozone effect, interference)
- DSA-negative ≠ safe (Consider non-HLA antibodies, technical false negatives)

PRACTICAL ALGORITHM – SUMMARY



★ **Key Takeaway:** Request DSA routinely and with any clinical trigger; monitor trends over time, and always combine with biopsy for diagnosis and management decisions.

DONOR-DERIVED CELL-FREE DNA (dd-cfDNA) IN CLINICAL PRACTICE

A non-invasive biomarker of allograft injury to guide transplant care

1. WHAT IS dd-cfDNA?



- Fragments of DNA released from injured donor allograft cells into recipient plasma
- Measured as % of total cfDNA or copies/mL plasma
- Reflects active graft injury from any cause

Examples of available assays:



2. CLINICAL USES



Early detection of rejection
Often precedes change in creatinine



Differentiate injury vs stable graft
Helps determine need for biopsy



Rule-out tool
High negative predictive value (NPV)



Risk stratification
Best when combined with DSA and clinical context

3. INTERPRETATION (KIDNEY TRANSPLANT)

dd-cfDNA (% of total cfDNA)

Interpretation

< 0.5%

Low probability of active rejection

0.5 – 1.0%

Indeterminate ("gray zone")

> 1.0%

Increased suspicion for rejection (esp. AMR)



Rising trend over time is more important than a single value

4. dd-cfDNA + DSA: HOW TO INTERPRET

dd-cfDNA	DSA	Interpretation	Suggested Action
Elevated (>1.0% or rising)	DSA Positive (de novo or persistent)	High risk of ANTIBODY-MEDIATED REJECTION (Active endothelial injury)	Biopsy recommended Treat if biopsy-proven
Elevated (>1.0% or rising)	DSA Negative	Graft injury from other causes (TCMR, BK, infection, ischemia, drug toxicity)	Biopsy recommended to identify cause
Normal / Low (<0.5%)	DSA Positive (de novo or persistent)	Possible SUBCLINICAL REJECTION (esp. AMR) Injury may be below detection threshold	Close monitoring Consider biopsy based on risk & clinical context
Normal / Low (<0.5%)	DSA Negative	Low probability of active rejection	Continue routine monitoring

5. WHEN TO USE dd-cfDNA



Routine surveillance
(e.g., 1, 3, 6, 12 months)



Any graft dysfunction
(creatinine ↑ or proteinuria)



New de novo DSA



Post-treatment monitoring
of rejection

6. MONITORING



- Stable patient: every 3–6 months
- High-risk / abnormal: every 1–2 months
- Focus on trend, not single value

7. LIMITATIONS



Not rejection-specific:
Infection (e.g., BK virus), ischemia, biopsy injury, other graft injuries



Early post-transplant levels may be elevated



Cannot distinguish AMR vs TCMR alone



Cost and accessibility limitations

8. KEY TAKEAWAY



dd-cfDNA is a **non-invasive biomarker** of allograft injury with **high negative predictive value**. It is best used **alongside DSA and clinical data** to guide biopsy decisions and optimize management of transplant recipients.



REFERENCES:

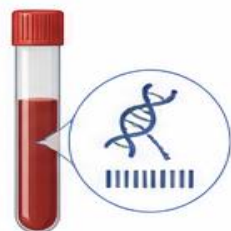
Bloom RD, Bromberg JS, Poggio ED, et al. Cell-Free DNA and Active Rejection in Kidney Allografts. *Circulation*. 2017;136(23):230–241.
Position Statement on Use of Donor-Derived Cell-Free DNA in Transplantation. *Am J Transplant*. 2023.
(*Kidney International* (2020) 98, 76–107).

American Society of Transplantation (AST).
Loupy A, Haas M, Solez K, et al. The Banff 2019 Kidney Meeting Report
(*Kidney International* (2020) 98, 76–107).

GENE EXPRESSION PROFILING (GEP) IN TRANSPLANTATION

A molecular "snapshot" of immune activity to guide transplant care

1. WHAT IS GEP?



- Measures mRNA expression patterns from peripheral blood or allograft biopsy tissue
- Reflects immune activation status (rejection vs quiescence)
- Uses multi-gene algorithms to generate a risk result

SOURCES

- Peripheral blood
- Allograft biopsy tissue

2. COMMON PLATFORMS / TESTS



AlloMap®
Heart transplant (blood test)

- Identifies low risk of acute cellular rejection



TruGraf®
Kidney transplant (blood test)

- Detects immune quiescence vs subclinical rejection



Tissue-based platforms
Molecular microscope / transcriptomics (e.g., MMDx) (biopsy tissue; research/advanced use)

3. CLINICAL USES



Non-invasive surveillance
Monitor immune status over time



Detect subclinical rejection
Identify immune activation before clinical dysfunction



Rule-out tool
High negative predictive value to reduce unnecessary biopsies



Works best in combination with DSA, dd-cfDNA and clinical data

4. INTERPRETATION (CONCEPTUAL)

GEP REPORT (Typical Output)

IMMUNE QUIESCENCE SIGNATURE

Low probability of rejection
(Stable / not-at-risk)



REJECTION / INJURY SIGNATURE

Increased risk of rejection
(At-risk / further evaluation needed)

Trend over time is more important than a single result

5. INTEGRATING GEP WITH OTHER BIOMARKERS

GEP RESULT	dd-cfDNA	DSA	INTERPRETATION	SUGGESTED ACTION
Normal (Quiescence)	Normal (<0.5%)	Negative	Very low risk of active rejection	Continue routine monitoring
Abnormal (Rejection signature)	Elevated (>1.0%)	Positive or Negative	Active injury likely	Biopsy recommended Treat if biopsy-proven
Normal (Quiescence)	Elevated (>1.0%)	Positive (de novo or persistent)	Possible early / subclinical AMR	Consider biopsy Close monitoring
Abnormal (Rejection signature)	Normal (<0.5%)	Negative	Immune activation without injury (e.g., early TCMR, viral, other)	Clinical correlation Repeat testing / Consider biopsy

GEP complements, but does not replace, clinical judgment and biopsy when indicated.

6. WHEN TO USE GEP



Routine surveillance (e.g., months 1, 3, 6, 12...)



Stable patient with clinical suspicion



Discordant DSA / dd-cfDNA results



To avoid biopsy in low-risk patients



Post-treatment monitoring

7. LIMITATIONS



Not rejection-specific (infection, inflammation, viral, effect)



Less validated than biopsy in kidney transplant



Variable performance in TCMR vs AMR



Platform-dependent algorithms



Cost and availability limitations

8. KEY TAKEAWAY



Gene expression profiling provides a non-invasive assessment of immune activity. It is best used to rule out rejection and reduce unnecessary biopsies, especially when combined with DSA and dd-cfDNA.

9. PRACTICAL ALGORITHM (How to Use GEP in Clinical Practice)



1. Identify Patient
Stable surveillance or clinical trigger



2. Order Tests
GEP + dd-cfDNA ± DSA (consider clinical context)



3. Interpret Together
Integrate GEP result with dd-cfDNA, DSA and clinical data



4. Decide Next Step
Biopsy if high risk / discordant
Monitor if low risk



5. Act & Reassess
Treat if biopsy-proven
Monitor trends over time

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cAMR DIAGNOSIS WITHOUT BIOPSY

A Practical Diagnostic Algorithm

KIDNEY TRANSPLANT RECIPIENT

Clinical trigger: ↑ Serum creatinine / proteinuria / ↓ eGFR / unexplained graft dysfunction

1

INITIAL RISK ASSESSMENT

Review history and baseline parameters

- Time from transplant
- Previous rejection episodes
- Non-adherence / under-immunosuppression
- Recent infections / comorbidities
- Immunologic risk (sensitization, high PRA)
- Trend of kidney function (stable vs progressive)

2

BASIC NON-INVASIVE SCREENING (ALL PATIENTS)



DSA TESTING

- Anti-HLA DSA
- Consider non-HLA (AT1R, ETAR, MICA)



DONOR-DERIVED CELL-FREE DNA (dd-cfDNA)

- e.g., AlloSure®, Prospera®



GENE EXPRESSION PROFILING (GEP)

- e.g., TruGraf®, kSORT®
- Detects immune activation signatures



ENDOTHELIAL BIOMARKERS

- CXCL9, CXCL10
- VCAM-1, E-selectin
- (± C4d signature)

3

INTEGRATE RESULTS

HIGH PROBABILITY OF cAMR

Any of the following:

- Strong DSA (high MFI) **PLUS**
- dd-cfDNA significantly ↑ (>1% or above assay cut-off)
- GEP with ABMR probability above threshold
- Endothelial markers markedly ↑ (CXCL9/10, VCAM-1)

LIKELY ACTIVE cAMR

(High non-invasive probability)

INTERMEDIATE PROBABILITY

Some of the following:

- Low/Moderate DSA or non-HLA antibodies
- dd-cfDNA mildly ↑ or borderline
- GEP borderline / inconclusive
- Mild elevation of endothelial markers

POSSIBLE cAMR

(Indeterminate risk)

LOW PROBABILITY

All of the following:

- No DSA / non-HLA Ab
- dd-cfDNA low (<0.5%)
- GEP not suggestive of rejection
- Endothelial markers normal or minimally ↑

LOW LIKELIHOOD OF cAMR

(Consider other causes)

CONFLICTING RESULTS

Discordant findings (e.g., DSA negative but dd-cfDNA high or GEP positive)

UNCERTAIN

(Needs further evaluation)

4

NEXT STEPS

- Consider this as probable active cAMR
- Optimize immunosuppression
- Treat if clinically indicated (per center protocol)
- **STRONGLY CONSIDER BIOPSY** to confirm & grade severity (Banff)

- Close clinical monitoring
- Repeat tests in 1–2 weeks
- Search for other causes (drug toxicity, infection, BK, obstruction)
- **CONSIDER BIOPSY** if persistent dysfunction or rising biomarkers

- Investigate non-immune causes (dehydration, drug toxicity, UTI, obstruction, BK, etc.)
- Routine monitoring
- Repeat panel if kidney function worsens

- Reassess clinically
- Repeat panel
- If uncertainty persists or kidney function declining → **PROCEED TO BIOPSY**

5

WHEN TO PROCEED DIRECTLY TO BIOPSY (Regardless of non-invasive results)



Rapid rise in creatinine (>20–30% within 1–2 weeks)



Nephrotic-range proteinuria or rapidly increasing proteinuria



Declining graft function with hemodynamic instability



New structural abnormality on imaging (if available)



High clinical suspicion of rejection despite negative tests

KEY POINTS



- No single non-invasive test can replace biopsy.
- Use a combination of tests to increase diagnostic confidence.
- Serial testing and trends are more informative than single values.
- Biopsy remains gold standard for definitive diagnosis, staging and prognosis.

ABBREVIATIONS

cAMR: chronic antibody-mediated rejection
 DSA: donor-specific antibody GEP: gene expression profiling
 dd-cfDNA: donor-derived cell-free DNA
 MFI: mean fluorescence intensity Ab: antibody

WRAP-UP: INTEGRATED NON-INVASIVE DIAGNOSIS OF cAMR

No single test is sufficient → Combine biomarkers + biopsy when needed

1. STEP 1: SCREEN (NON-INVASIVE BIOMARKERS)

DSA

Immunologic trigger



- Detect de novo DSA
- Monitor trend
- Class II (DQ) highest risk

dd-cfDNA

Graft injury



- Reflects allograft injury
- Rising levels increase risk

GEP

Immune activation



- Gene expression signature
- Detects immune activation

2. STEP 2: INTERPRET PATTERN (TRIAD APPROACH)

DSA	dd-cfDNA	GEP	INTERPRETATION	SUGGESTED ACTION
Positive (de novo or persistent)	Elevated (>1.0% or rising)	Abnormal (Rejection signature)	HIGH PROBABILITY AMR Active antibody-mediated injury likely	Biopsy recommended
Positive (de novo or persistent)	Normal (<0.5%)	Normal (Quiescence signature)	POSSIBLE SUBCLINICAL AMR No current injury but immune risk present	Monitor closely Consider biopsy (based on risk)
Negative	Elevated (>1.0% or rising)	Abnormal (Rejection signature)	NON-DSA INJURY Consider TCMR, infection, drug toxicity, ischemia, etc.	Biopsy recommended
Negative	Normal (<0.5%)	Normal (Quiescence signature)	LOW RISK Very low probability of active rejection	No biopsy Continue routine monitoring

3. STEP 3: CONFIRM

KIDNEY BIOPSY = GOLD STANDARD



- Microvascular inflammation
- Transplant glomerulopathy (TG)
- C4d staining (±)
- Plasma cell infiltration
- Chronic changes

Histology + clinical context = Definitive diagnosis

4. STEP 4: RISK STRATIFICATION



High risk of cAMR if:

- De novo DSA (especially DQ)
- Rising dd-cfDNA (trend)
- Persistent abnormal GEP
- Complement-binding DSA (C1q+, C3d+)
- Prior rejection or non-adherence

5. DIAGNOSTIC PITFALLS

- ✗ Treating biomarkers without biopsy
- ✗ Ignoring discordant results
- ✗ Assuming DSA-negative = no AMR
- ✗ Relying on a single value (trend is more important)

6. FINAL TAKEAWAY



Early detection of cAMR requires an integrated approach:
DSA (trigger) + dd-cfDNA (injury) + GEP (immune activity)
 → **Biopsy confirmation**

7. PRACTICAL ALGORITHM FOR DIAGNOSIS OF cAMR

